Biochemical and Biophysical Research Communications xxx (2012) xxx-xxx

Contents lists available at SciVerse ScienceDirect



Biochemical and Biophysical Research Communications



21

22

23

24

25

26

27

28

29

30 31

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

journal homepage: www.elsevier.com/locate/ybbrc

² The adult polyglucosan body disease mutation *GBE1* c.1076A>C occurs at high ³ frequency in persons of Ashkenazi Jewish background

⁴ Q1 Abrar Hussain^a, Joy Armistead^a, Lara Gushulak^a, Christa Kruck^a, Steven Pind^a, Barbara Triggs-Raine^a, ⁵ Marvin R. Natowicz^{b,*}

⁶ ^a Department of Biochemistry and Medical Genetics, University of Manitoba, Winnipeg, Canada

^b Pathology and Laboratory Medicine Institutes and Genomic Medicine, Neurological and Pediatrics Institutes, Cleveland Clinic, Cleveland, OH 44195, USA

ARTICLE INFO

20 12

8

Article history:
Received 10 August 2012

14 Available online xxxx

- 15 Q2 Keywords:
- 16 Glycogen branching enzyme
- 17 Polyglucosan body disease
- 18 Ashkenazi Jewish

ABSTRACT

Mutations of the glycogen branching enzyme gene, *GBE1*, result in glycogen storage disease (GSD) type IV, an autosomal recessive disorder having multiple clinical forms. One mutant allele of this gene, *GBE1* c.1076A>C, has been reported in Ashkenazi Jewish cases of an adult-onset form of GSD type IV, adult polyglucosan body disease (APBD), but no epidemiological analyses of this mutation have been performed. We report here the first epidemiological study of this mutation in persons of Ashkenazi Jewish background and find that this mutation has a gene frequency of 1 in 34.5 (95% CI: 0.0512–0.0145), similar to the frequency of the common mutation causing Tay-Sachs disease among Ashkenazi Jews. This finding reveals APBD to be another monogenic disorder that occurs with increased frequency in persons of Ashkenazi Jewish ancestry.

© 2012 Published by Elsevier Inc.

32

33 1. Introduction

The glycogen biosynthetic enzyme gene GBE1 encodes the gly-34 cogen branching enzyme (EC 2.4.1.15), an enzyme that catalyzes 35 the transfer of alpha-1,4 linked glucosyl units from the outer end 36 of a glycogen chain to an alpha-1,6 position on the same or nearby 37 oligosaccharide chain. This enzyme activity results in the branch-38 ing of high molecular weight glycogen molecules and thereby 39 enables the packing of a large number of glycosyl units into a rel-40 atively soluble spherical molecule [1]. 41

The absence or defective function of glycogen branching en-42 zyme results in one of several clinical forms of glycogen storage 43 disease type IV (GSD type IV; OMIM #232500) in humans and in 44 mouse and other animal models [1-3]. Despite marked heteroge-45 neity in age of onset and natural history, all clinical forms of GSD 46 47 type IV are associated with an accumulation of structurally 48 abnormal glycogen in tissues and diminished or absent glycogen branching enzyme activity. In most instances where mutation 49 analysis has been carried out, two pathologic DNA sequence vari-50 ants of the GBE1 gene have been found, consistent with autosomal 51 recessive inheritance. 52

GSD type IV is a panethnic disorder. One form of GSD type IV, an
adult-onset form termed adult polyglucosan body disease (APBD;
OMIM #263570), occurs in persons of diverse backgrounds but

E-mail address: natowim@ccf.org (M.R. Natowicz).

0006-291X/\$ - see front matter © 2012 Published by Elsevier Inc. http://dx.doi.org/10.1016/j.bbrc.2012.08.089 many of the reported cases are of Ashkenazi Jewish background [4]. It, like other forms of GSD type IV, is due to reduced glycogen branching enzyme activity [5].

The mutations of *GBE1* reported in persons having APBD are heterogeneous [6–9], although all persons of Ashkenazi Jewish background reported to date have at least one copy of the c.1076A>C mutation. This raises the question whether there is an increased frequency of c.1076A>C among individuals of Ashkenazi Jewish background. No population-based studies from which to make accurate epidemiologic assessments have yet been done.

The epidemiology of numerous monogenic and multifactorially determined conditions has been studied in detail in persons of Ashkenazi Jewish background, with some disorders and their associated mutations occurring at increased levels relative to many other populations [10,11]. We report here the first epidemiologic study of the mutation frequency of the ABPD-associated *GBE1* mutation c.1076A>C in a large Ashkenazi Jewish cohort.

2. Materials and methods

2.1. Clinical samples

The samples used in this study were de-identified leukocyte 75 pellets previously used for clinical carrier screening for Tay-Sachs 76 disease. All individuals had provided informed consent for carrier 77 screening and had stated that all four grandparents were of 78 Ashkenazi Jewish heritage. Genomic DNA was extracted from the 79 leukocyte pellets by standard methods [12]. 80

Please cite this article in press as: A. Hussain et al., The adult polyglucosan body disease mutation *GBE1* c.1076A>C occurs at high frequency in persons of Ashkenazi Jewish background, Biochem. Biophys. Res. Commun. (2012), http://dx.doi.org/10.1016/j.bbrc.2012.08.089

^{*} Corresponding author. Address: Cleveland Clinic, LL-3, 9500 Euclid Avenue, Cleveland, OH 44195, USA. Fax: +1 216 445 9212.

ARTICLE IN PRESS

A. Hussain et al. / Biochemical and Biophysical Research Communications xxx (2012) xxx-xxx



Fig. 1. Detection strategy for *GBE1* c.1076A>C. DNA was amplified with primers that create an Hnfl site only in the presence of the c.1076A>C mutation. PCR products were digested with Hnfl and separated on a 2% agarose gel. Lanes: 1-Molecular Weight Marker, 2-Control PCR Product, 3-Carrier PCR Product. An asterisk indicates the new 123 bp DNA fragment created only in the presence of the mutation. The 19 bp fragment is run off the bottom of the gel.

81 2.2. Detection of the GBE1 c.1076A>C mutation

The region harboring the GBE1 c.1076A>C mutation (NM_ 82 83 000158) was PCR-amplified with forward primer 5'-tgggatagcagattgtttgact-3' and reverse primer 5'-taagaacattaagtaccagtga-3' 84 85 using an annealing temperature of 55 °C. All other PCR conditions 86 are as described previously [13]. A site-directed C to A substitution in the forward primer (underlined), resulted in an Hnf1 restriction 87 enzyme site that created a 123 bp fragment only in the presence of 88 89 the 1076A>C mutation. The 142 bp PCR product and 123/19 bp 90 restriction products were separated on a 2% agarose gel or an 8% 91 acrylamide gel and visualized by ethidium bromide staining (Fig. 1).

92 2.3. Detection of the HEXA c.1278insTATC mutation

93 For the detection of the c.1278insTATC mutation in exon 11 of 94 HEXA, a 387 bp PCR-product was generated with forward primer 95 5'-tcccatgttgcctgtgtatg-3' and reverse primer 5'-cctcaccccagctaag 96 ttgt-3'. The mutation was detected by the presence of an additional 97 elution peak using denaturing high performance liquid chromatog-98 raphy (dHPLC). Samples were separated on a Transgenomics 99 Wave[®] 3500 system using a manufacturer supplied DNASep[®] col-100 umn and Wave optimized buffers (Transition Technologies, Inc., 101 Toronto). Separations were performed at an oven temperature of 102 58.4 °C using a gradient of 56.8-65.8% buffer B. Samples with an 103 additional peak by dHPLC analysis were confirmed to have the 104 c.1278insTATC mutation by heteroduplex analysis as described 105 previously [14].

106 **3. Results and discussion**

DNA was successfully analyzed for the presence/absence of 107 108 GBE1 c.1076A>C in samples from 380 persons who reported that all four grandparents were of Ashkenazi Jewish background. Eleven 109 samples had this mutation (Fig. 1), resulting in a heterozygote 110 frequency of 1 in 34.5 persons (95% CI: 0.0512-0.0145), an unex-111 112 pectedly high carrier frequency that approximates the carrier fre-113 quencies of the most common deleterious mutations for several 114 disorders that more commonly occur in persons of Ashkenazi Jew-115 ish background [10,11].

A possible explanation for the high heterozygote frequency of c.1076A>C in *GBE1* noted above is a biased or otherwise unusual

representation of persons of Ashkenazi Jewish background in the 118 cohort studied here. In consideration of this possibility, we then 119 assessed the frequency of the most common recessive mutation 120 for Tay-Sachs disease in persons of Jewish background, the HEXA 121 c.1278insTATC mutation, in the same cohort. DNA was successfully 122 analyzed for the presence/absence of the HEXA c.1278insTATC al-123 lele in 402 samples. Fourteen samples had this mutation, resulting 124 in a heterozygote frequency of 1 in 28.7 (95% CI: 0.0577-0.0192), 125 similar to the heterozygote frequency of HEXA c.1278insTATC re-126 ported for American Ashkenazi Jews as well as Ashkenazi Jews in 127 other countries [15-20]. This result, in turn, makes it unlikely that 128 the high frequency of GBE1 c.1076A>C observed in this study is due 129 to a sampling bias. 130

The finding of a high frequency of a disease-associated allele is not unique in this population. Much work has been done on the population genetics of persons of Jewish background, especially Ashkenazi Jews [21–23]. Among the mutant alleles for serious disorders that occur at high frequencies in persons of Ashkenazi Jewish background, only specific mutant alleles for Gaucher disease (*GBA* p.409S), Tay-Sachs disease (*HEXA* c.1278insTATC) and familial dysautonomia (*IKBKAP* c.2204+6T>C) occur at similar or higher frequencies than the *GBE1* c.1076A>C heterozygote frequency reported here [10,11]. The determination of the origin and subsequent population history of the *GBE1* c.1076A>C mutation and how this relates to its high prevalence in persons of Ashkenazi Jewish background is an important question that arises from this work.

Sequencing of all exons and intron/exon junctions typically re-145 veals two allelic mutations of the GBE1 locus in Ashkenazi Jews with 146 ABPD, usually homozygosity of c.1076A>C. There are, however, sev-147 eral instances of clinically, neuroradiologically and biopsy-proven 148 APBD in Ashkenazi Jewish individuals where mutation analysis re-149 vealed only a single copy of that mutant allele and an apparent ab-150 sence of another allelic mutation ([24], M. Natowicz, personal 151 observations), as well as a report of a non-Jewish individual with 152 clinically-definite APBD and only a single detected mutant GBE1 153 mutation [9]. These cases, in turn, raise several interesting possibil-154 ities: (1) that a small percentage of individuals with APBD are man-155 ifesting heterozygotes of a single mutation of *GBE1*; (2) that there is 156 an undetected mutation of the other allele of GBE1 in these individ-157 uals, such as a promoter mutation in trans to the detected mutant 158 allele; or (3) that these individuals harbor a mutation of another 159 gene that acts synergistically with the single mutated allele. Thus 160 far, only coding region or splice junction mutations have been re-161 ported for the GBE1 gene. Synergistic heterozygosity as the basis 162 for a subset of cases of APBD is without precedent and, if present, 163 would add to our understanding of the complexity of glycogen 164 metabolism. Two instances of possible synergistic heterozygosity 165 related to heterozygosity for myophosphorylase, another enzyme 166 of glycogen metabolism, have been reported [25]. 167

APBD now appears to be another genetic condition prevalent in individuals of Ashkenazi Jewish background. Insofar as APBD is clinically heterogeneous in its initial presentation and natural history, it is often difficult to diagnose and the correct diagnosis can be delayed for many years; affected individuals are often misdiagnosed as having multiple sclerosis or prostatic hyperplasia [4]. The findings of this study have particular relevance for persons of Ashkenazi Jewish background for whom APBD might be a clinical consideration. This work also raises interesting questions regarding the population history of this mutation and its impacts on glycogen metabolism.

Acknowledgments

179

180

181

We are grateful to the Louis and Susan Coddon Family Foundation for support of this work.

Please cite this article in press as: A. Hussain et al., The adult polyglucosan body disease mutation *GBE1* c.1076A>C occurs at high frequency in persons of Ashkenazi Jewish background, Biochem. Biophys. Res. Commun. (2012), http://dx.doi.org/10.1016/j.bbrc.2012.08.089

2

176

177

178

168

169

170

171

131

132

133

134

135

136

137

138

139

140

141

142

143

144

ARTICLE IN PRESS

A. Hussain et al./Biochemical and Biophysical Research Communications xxx (2012) xxx-xxx

182 References

188

189

191

192

193

194

195

196

197

198

199

200

201

202

203

204

205

206

207

208

212

213

214

- 183 [1] P.S. Kishnani, D. Koeberl, Y.-T. Chen, Glycogen storage diseases, in: D. Valle, A.L. 184 Beaudet, B. Vogelstein, K.W. Kinzler, S.E. Antonarakis, A. Ballabio (Eds.), 185 Scriver's Online Metabolic and Molecular Basis of Inherited Disease, McGraw-186 Hill, New York, 2009 (Chapter 71). 187
 - [2] Y.-C. Lee, C.-J. Chang, D. Bali, et al., Glycogen-branching enzyme deficiency leads to abnormal cardiac development: novel insights into glycogen storage disease IV. Hum. Mol. Genet. 20 (2011) 455-465.
- 190 [3] H.O. Akman, T. Sheiko, S.K.H. Tay, et al., Generation of a novel mouse model that recapitulates early and adult onset glycogenosis type IV, Hum. Mol. Genet. 20 (2011) 4430-4439
 - [4] C.J. Klein, Adult polyglucosan body disease, in: R.A. Pagon, T.D. Bird, C.R. Dolan, et al. (Eds.). GeneReviews™ [Internet]. Seattle, WA. 2009.
 - [5] A. Lossos, V. Barash, D. Soffer, et al., Hereditary branching enzyme dysfunction in adult polyglucosan body disease; a possible metabolic cause in two patients. Annu. Neurol. 30 (1991) 655-661.
 - [6] A. Lossos, Z. Meiner, V. Barash, et al., Adult polyglucosan body disease in Ashkenazi Jewish patients carrying the Tyr³²⁹Ser mutation in the glycogenbranching enzyme gene, Annu. Neurol. 44 (1998) 867–872.
 - F. Ziemssen, E. Sindern, J.M. Schroder, et al., Novel missense mutations in the [7] glycogen-branching enzyme gene in adult polyglucosan body disease, Annu. Neurol. 47 (2000) 536-540.
 - [8] E. Sindern, F. Ziemssen, T. Ziemssen, et al., Adult polyglucosan body disease: a postmortem correlation study, Neurology 61 (2003) 263-265.
 - [9] R. Massa, C. Bruno, A. Martorana, et al., Adult polyglucosan body disease: proton magnetic resonance spectroscopy of the brain and novel mutation in the GBE1 gene, Muscle Nerve 37 (2008) 530-536.
- 209 [10] J. Charrow, Ashkenazi Jewish genetic disorders, Fam. Cancer 3 (2004) 201–206. 210 [11] S.J. Gross, B.A. Pletcher, K.G. Monaghan, Carrier screening in individuals of 211
 - Ashkenazi Jewish descent, Genet. Med. 10 (2008) 54-56. [12] S. Pind, E. Slominski, J. Mauthe, et al., V490M, a common mutation in 3-
 - phosphoglycerate dehydrogenase deficiency, causes enzyme deficiency by decreasing the yield of mature enzyme, J. Biol. Chem. 277 (2002) 7136–7143. J. Armistead, S. Kharker, B. Meyer, et al., Mutation of a gene essential for [13]
- 215 216 ribosome biogenesis, EMG1, causes Bowen-Conradi syndrome, Am. J. Hum. 217 Genet. 84 (2009) 728-739.

- [14] B.L. Triggs-Raine, R.A. Gravel, Diagnostic heteroduplexes: simple detection of carriers of a 4-bp insertion mutation in Tay-Sachs disease, Am. J. Hum. Genet. 46 (1990) 183-184.
- [15] M. Kaback, J. Lim-Steele, D. Dabholkar, et al., Tay-Sachs disease carrier screening, prenatal diagnosis, and the molecular era: an international perspective, 1970-1993, JAMA 270 (1993) 2307-2315.
- [16] J.M. DeMarchi, C.T. Caskey, C.S. Richards, Population-specific screening by mutation analysis for diseases frequent in Ashkenazi Jews, Hum. Mutat. 8 (1996) 116–125.
- [17] G. Bach, J. Tomczak, N. Risch, J. Eckstein, Tay-Sachs screening in the Jewish Ashkenazi populations: DNA testing is the preferred procedure, Am J. Med. Genet. 99 (2001) 70-75.
- [18] R. Rozenberg, L. da Veiga Periera, The frequency of Tay-Sachs disease causing mutations in the Brazilian Jewish population justifies a carrier screening program, Sao Paulo Med. J. 119 (2001) 146-149.
- [19] S.A. Scott, L. Edelman, L. Liu, et al., Experience with carrier screening and prenatal diagnosis for 16 Ashkenazi Jewish genetic diseases, Hum. Mutat. 31 (2010) 1240-1250.
- [20] R. Lew, L. Burnett, A. Proos, Tay-Sachs disease preconception screening in Australia: self-knowledge of being an Ashkenazi Jew predicts carrier state better than does ancestral origin, although there is an increased risk for c.1421+1G>C mutation in individuals with South African heritage, J. Commun. Genet. 2 (2011) 201-209.
- G. Atzmon, L. Hao, I. Pe'er, et al., Abraham's children in the genome era: major Jewish diaspora populations comprise distinct genetic clusters with shared Middle Eastern ancestry, Am. J. Hum. Genet. 86 (2010) 850-859.
- [22] D.M. Behar, B. Yunusbayev, M. Metspalu, et al., The genome-wide structure of the Jewish people, Nature 466 (2010) 238-243.
- S. Guha, J.A. Rosenfeld, A.K. Malhotra, et al., Implications for health and disease in the genetic signature of the Ashkenazi Jewish population, Genome Biol. 13 (2012) R2.
- [24] E.E. Ubogu, S.T.K. Hong, H.O. Akman, et al., Adult polyglucosan body disease: a case report of a manifesting heterozygote, Muscle Nerve 32 (2005) 675-681.
- [25] J. Vockley, P. Rinaldo, M.J. Bennett, et al., Synergistic heterozygosity: disease resulting from multiple partial defects in one or more metabolic pathways, Mol. Genet. Metab. 71 (2000) 10-18.

252 253 254

249

250

251

3

Please cite this article in press as: A. Hussain et al., The adult polyglucosan body disease mutation GBE1 c.1076A>C occurs at high frequency in persons of Ashkenazi Jewish background, Biochem. Biophys. Res. Commun. (2012), http://dx.doi.org/10.1016/j.bbrc.2012.08.089